

Client: American Mold Experts
C/O: Mr Bill Nicoll, cmi
Re: Edwards, Post Test

Date of Sampling: 11-06-2018
Date of Receipt: 11-07-2018
Date of Report: 11-07-2018

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	A1: 2nd Floor hall, return			A2: Living room, center		
Comments (see below)	None			None		
Lab ID-Version‡:	9618352-1			9618353-1		
Analysis Date:	11/07/2018			11/07/2018		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Ascospores				2	100	27
Basidiospores	10	100	130	10	100	130
Chaetomium						
Cladosporium	3	100	40	5	100	67
Curvularia						
Epicoccum						
Fusarium						
Myrothecium						
Nigrospora				1	100	13
Other colorless						
Penicillium/Aspergillus types†	4	100	53	10	100	130
Pithomyces						
Rusts				2	100	27
Smuts, Periconia, Myxomycetes				1	100	13
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)††	2+			3+		
Hyphal fragments/m3	40			53		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	2+			2+		
Sample volume (liters)	75			75		
§ TOTAL SPORES/m3			230			410

Comments:

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

†† Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m³ divided by the raw count, expressed in spores/m³. The limit of detection is the analytical sensitivity (in spores/m³) multiplied by the sample volume (in liters) divided by 1000 liters.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

§ Total Spores/m³ has been rounded to two significant figures to reflect analytical precision.

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SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	A3: Basement, center		
Comments (see below)	None		
Lab ID-Version‡:	9618354-1		
Analysis Date:	11/07/2018		
	raw ct.	% read	spores/m3
Ascospores			
Basidiospores	10	100	130
Chaetomium			
Cladosporium	3	100	40
Curvularia			
Epicoccum			
Fusarium			
Myrothecium			
Nigrospora			
Other colorless			
Penicillium/Aspergillus types†	17	100	230
Pithomyces			
Rusts			
Smuts, Periconia, Myxomycetes			
Stachybotrys			
Stemphylium			
Torula			
Ulocladium			
Zygomycetes			
Background debris (1-4+)††	1+		
Hyphal fragments/m3	27		
Pollen/m3	< 13		
Skin cells (1-4+)	1+		
Sample volume (liters)	75		
§ TOTAL SPORES/m3			400

Comments:

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DIRECT MICROSCOPIC EXAMINATION REPORT

Background Debris and/or Description	Miscellaneous Spores Present*	MOLD GROWTH: Molds seen with underlying mycelial and/or sporulating structures†	Other Comments††	General Impression
Lab ID-Version‡: 9618349-1, Analysis Date: 11/07/2018: Swab sample S1: Kitchen, wall cavity				
Light	Very few	None	None	Normal trapping
Lab ID-Version: 9618350-1, Analysis Date: 11/07/2018: Swab sample S2: Master bathroom, above shower				
Light	Very few	None	None	Normal trapping
Lab ID-Version: 9618351-1, Analysis Date: 11/07/2018: Swab sample S3: Crawl, joist				
Light	Very few	None	None	Normal trapping

* Indicative of normal conditions, i.e. seen on surfaces everywhere. Includes basidiospores (mushroom spores), myxomycetes, plant pathogens such as ascospores, rusts and smuts, and a mix of saprophytic genera with no particular spore type predominating. Distribution of spore types seen mirrors that usually seen outdoors.

† Quantities of molds seen growing are listed in the MOLD GROWTH column and are graded <1+ to 4+, with 4+ denoting the highest numbers.

†† Some comments may refer to the following: Most surfaces collect a mix of spores which are normally present in the outdoor environment. At times it is possible to note a skewing of the distribution of spore types, and also to note "marker" genera which may indicate indoor mold growth. Marker genera are those spore types which are present normally in very small numbers, but which multiply indoors when conditions are favorable for growth.

‡ A "Version" indicated by "-x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".
 The limit of detection is < 1+ when mold growth is detected.