EMLab P&K
3929 Old Lee Highway, Suite 91C, Fairfax, VA 22030
(866) 871-1984 Fax (856) 334-1040 www.emlab.com

Client: American Mold Experts
C/O: Mr Bill Nicoll, cmi
Re: Henderson, Pre Test

Date of Sampling: 04-03-2019
Date of Receipt: 05-01-2019
Date of Report: 05-01-2019

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	A1: Music room air return			A2: Basement air return		
Comments (see below)	None			None		
Lab ID-Version‡:	10199823-1			10199824-1		
Analysis Date:	05/01/2019			05/01/2019		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria	2	100	27			
Ascospores	1	25	53	1	25	53
Basidiospores	3	25	160	7	25	370
Chaetomium						
Cladosporium	4	25	210	1	25	53
Fusarium						
Myrothecium						
Nigrospora						
Other brown	2	100	27			
Other colorless						
Penicillium/Aspergillus types†	3	25	160	9	25	480
Pithomyces						
Rusts						
Smuts, Periconia, Myxomycetes	1	100	13			
Stachybotrys						
Stemphylium						
Torula	2	100	27			
Trichocladium	1	100	13			
Ulocladium						
Zygomycetes						
Background debris (1-4+)††	2+			3+		
Hyphal fragments/m3	67			160		
Pollen/m3	27			< 13		
Skin cells (1-4+)	1+			1+		
Sample volume (liters)	75			75		
§ TOTAL SPORES/m3			690			960

Comments:

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.
† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium, Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.
††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m^3 divided by the raw count, expressed in spores/m^3. The limit of detection is the analytical sensitivity (in spores/m^3) multiplied by the sample volume (in liters) divided by 1000 liters.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.
‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".
§ Total Spores/m3 has been rounded to two significant figures to reflect analytical precision.

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SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	A3: Upstairs air return		
Comments (see below)	None		
Lab ID-Version‡:	10199825-1		
Analysis Date:	05/01/2019		
	raw ct.	% read	spores/m3
Alternaria			
Ascospores	1	25	53
Basidiospores	1	25	53
Chaetomium			
Cladosporium	1	25	53
Fusarium			
Myrothecium			
Nigrospora			
Other brown			
Other colorless			
Penicillium/Aspergillus types†	3	25	160
Pithomyces			
Rusts			
Smuts, Periconia, Myxomycetes	1	100	13
Stachybotrys			
Stemphylium			
Torula			
Trichocladium			
Ulocladium			
Zygomycetes			
Background debris (1-4+)††	3+		
Hyphal fragments/m3	13		
Pollen/m3	27		
Skin cells (1-4+)	1+		
Sample volume (liters)	75		
§ TOTAL SPORES/m3			330

Comments:

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.
† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium, Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.
††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

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DIRECT MICROSCOPIC EXAMINATION REPORT

Background Debris and/or Description	Miscellaneous Spores Present*	MOLD GROWTH: Molds seen with underlying mycelial and/or sporulating structures†	Other Comments††	General Impression
Lab ID-Version‡: 10199821-1, Analysis Date: 05/01/2019: Swab sample S1: Master bath				
Scant	Very few	None	None	Normal trapping
Lab ID-Version: 10199822-1, Analysis Date: 05/01/2019: Swab sample S2: Sump box				
Scant	None	None	None	No mold spores detected

* Indicative of normal conditions, i.e. seen on surfaces everywhere. Includes basidiospores (mushroom spores), myxomycetes, plant pathogens such as ascospores, rusts and smuts, and a mix of saprophytic genera with no particular spore type predominating. Distribution of spore types seen mirrors that usually seen outdoors.

† Quantities of molds seen growing are listed in the MOLD GROWTH column and are graded <1+ to 4+, with 4+ denoting the highest numbers.

†† Some comments may refer to the following: Most surfaces collect a mix of spores which are normally present in the outdoor environment. At times it is possible to note a skewing of the distribution of spore types, and also to note "marker" genera which may indicate indoor mold growth. Marker genera are those spore types which are present normally in very small numbers, but which multiply indoors when conditions are favorable for growth.

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".
The limit of detection is < 1+ when mold growth is detected.